

Empedobacter falsenii: a rare non-fermenter causing urinary tract infection in a child with bladder cancer

This article was published in the following Scient Open Access Journal:

SOA: Clinical Medical Cases, Reports & Reviews

Received July 17, 2017; Accepted August 10, 2017; Published August 17, 2017

Kamran Zaman, Parakriti Gupta, Varpreet Kaur, Balvinder Mohan and Neelam Taneja*

Postgraduate Institute of Medical Education and Research, Chandigarh, India

Abstract

Empedobacter falsenii is a rarely encountered gram-negative non-fermenting (NF) bacterium. It is most often misidentified due to limitations of conventional culture based identification methods and the true significance of its isolation remains obscure. However, in present times, the identification of NFs has become quick and reliable with the use of matrix-assisted laser desorption ionization time-of-flight mass spectrometry (MALDI-TOF). *E. falsenii*, which was formerly known as *Wautersiella falsenii*, is the only species of the genus. There are no definite guidelines for the antibiotic susceptibility testing and empirical therapy for this multidrug resistant agent. Till date in the world literature, only one case of urinary tract infection (UTI) by *E. falsenii* has been reported in a one-year-old child with pyelonephritis. Here, we report the first case of *E. falsenii* associated UTI from India which was identified using MALDI-TOF.

Keywords: *Empedobacter falsenii*, Non-fermenter, MALDI-TOF, Urinary Tract Infection, India

Introduction

The members of family *Enterobacteriaceae* are the most common etiological agents implicated in urinary tract infections (UTI), followed by gram-positive organisms such as *Staphylococci* and *Enterococci* [1]. UTI due to non-fermenters (NFs) have also seen an upsurge over the past few years. *Pseudomonas* holds a significant place in this group, due to easy identification by virtue of its pigment and odour, followed by *Acinetobacter* spp. Other rare NFs which have been reported to cause UTI include *Alcaligenes*, *Flavobacter*, *Oligella*, *Flavimonas*, *Agrobacter*, *Weeksiella*, *Chryseobacterium*, *Achromobacter* and *Elizabethkingia* [2,3]. Earlier, identification of these NF was labor-intensive and technically demanding which led to their misidentification as contaminants or isolates of doubtful significance. However, the identification of these organisms has now become reliable and rapid with the advent of rapid automatic identification methods like MALDI-TOF. Here, we report a case of UTI in a child caused by *Empedobacter falsenii*, a rare non fermenting, multidrug resistant NF, which was identified by MALDI-TOF. *Empedobacter falsenii*, belongs to family *Flavobacteriaceae*, which produces yellow colored pigment. There are no definite guidelines for the antibiotic susceptibility testing and treatment for this organism.

Case Report

A 5-year-old boy presented with ailments of fever, burning micturition while receiving radiotherapy for management of bladder cancer (rhabdomyosarcoma). A midstream urine sample was subjected to culture sensitivity to rule out urinary tract infection (UTI). Wet mount microscopic examination of uncentrifuged urine revealed 2-3 RBCs, abundant pus cells and bacteria per high power field. Semi-quantitative culture of the urine sample done on cysteine lysine electrolyte deficient (CLED) agar yielded significant bacterial growth (colony count $>10^5$ CFU/mL) of a yellow pigment-producing bacterium. The colonies were pale yellow, translucent, with entire edges on CLED medium and the organism could grow on MacConkey agar plate showing pale, yellow pigment producing translucent colonies. Figure 1, Gram stain of the isolate showed noncapsulated, nonsporing gram-negative bacilli. On conventional biochemical tests, it was catalase positive, oxidase positive, non-motile, produced indole and was negative for amino acid (lysine, ornithine, and arginine) decarboxylase, urease, and citrate utilization. This isolate was confirmed as *Empedobacter falsenii*, by matrix-assisted laser

*Corresponding author: Neelam Taneja, Postgraduate Institute of Medical Education and Research, Chandigarh, India, Tel: 0172-2755163, Email: drneelamgpi@yahoo.com

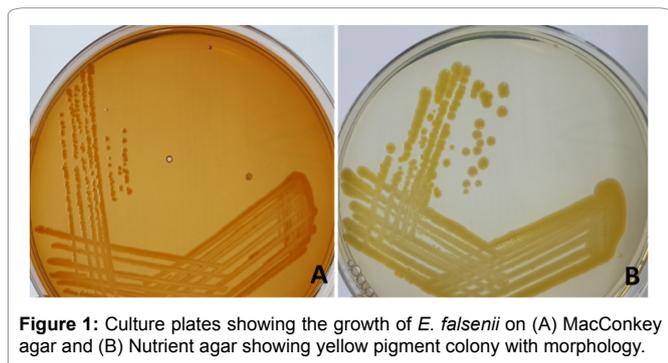


Figure 1: Culture plates showing the growth of *E. falsenii* on (A) MacConkey agar and (B) Nutrient agar showing yellow pigment colony with morphology.

desorption ionization time of flight mass spectrometry (MALDI-TOF, Bruker Daltonics, Bremen, Germany) with a score of 2.1. A repeat urine culture with all aseptic measures was advised to rule out any chance of contamination. The repeat urine culture also grew the same organism which was again confirmed by MALDI-TOF. Antimicrobial susceptibility was performed using the Kirby-Bauer disc diffusion method for: ampicillin (10µg), ampicillin-sulbactam (10µg), meropenem (10µg), ceftazidime (30µg), amikacin (30µg), cotrimoxazole (25µg), ciprofloxacin (5µg), gentamicin (10µg), imipenem (10µg), piperacillin-tazobactam (100/10µg), ceftriaxone (30µg), ampicillin (10µg) and colistin (300U) using previously published studies for reference due to the non-availability of CLSI guidelines [4,5]. The isolate showed sensitivity to gentamicin, amikacin, ceftriaxone and ceftazidime. The patient was started on intravenous amikacin and responded well to the treatment. The follow-up urine culture repeated after 2 weeks of therapy was sterile.

Discussion

E. falsenii is a gram-negative bacillus, belonging to family *Flavobacteriaceae*, which produces yellow colored pigment, grows aerobically at 37° C, and is catalase and oxidase positive. Indole is produced but the organism doesn't reduce nitrates and utilize citrate. It has weak-to-strong gelatinize activity. *Empedobacter* usually produces acid from glucose and maltose and comprises of two genomovars 1 and 2, which are phenotypically very similar [6,7]. Not a lot is known about epidemiology, clinical correlation and antimicrobial susceptibility of *Empedobacter*. The first clinical isolation of *Empedobacter* was described in a one-year-old child with complicated UTI in Netherlands in 2012. The organism was identified using MALDI-TOF and confirmed using 16s DNA PCR-sequencing [1]. Another case of *E. falsenii* was described by Traglia GM *et al*, in an 18-year-old female, who presented with cervical abscess associated with otitis media. The organism was identified by MALDI-TOF and confirmed by whole genome sequencing [2]. Recently in 2016, it has been isolated from a respiratory sample of a thirty two year old immunocompromised male having leukemia and was identified by MALDI-TOF [8]. The first two isolates were sensitive to ciprofloxacin, cefepime, gentamicin, cotrimoxazole and amikacin. But the isolate from the respiratory sample of leukemia patient showed resistance to many antibiotics like amikacin, ampicillin-clavulanic acid, ampicillin-sulbactam, cefotaxime, ceftazidime, doripenem, gentamicin, imipenem and piperacillin-tazobactam and it was sensitive only to cefepime and cotrimoxazole. Our isolate was susceptible to gentamicin, amikacin, ceftriaxone, ceftazidime was intermediately susceptible to piperacillin-tazobactam,

ciprofloxacin, imipenem and meropenem and was resistant to ampicillin, ampicillin-sulbactam and colistin. It's interesting to observe that previous isolates of other studies showed resistance to cephalosporins, whereas our isolate was sensitive to both ceftazidime and ceftriaxone. Exposure to antibiotics varies in different settings across the geographical regions. However, the exact reasons can better be evaluated by carrying out further studies to understand the resistance mechanisms that the organism adopts.

Empedobacter has earlier been reported from a patient with cystic fibrosis and prosthetic joint infection by 16s PCR. No significant clinical association was established as the organism was not isolated on culture [9]. The organism can also exist in rodent skin, soil, pollution sediments, machining facilities and even hospital carpets [10,11]. This implies that despite isolation, establishing a cause-effect relationship is difficult.

Conclusion

We report for the first time from India, *E. falsenii* causing UTI in a child with bladder cancer. Till date, there is only one other case report of *Empedobacter* causing UTI, which was seen in a one-year-old girl with pyelonephritis. As both these patients had an underlying illness, it can be hypothesized that prior instrumentation or previous surgery on genitourinary tract can predispose to such infections. The present report also emphasizes the significance of MALDI-TOF in the identification of these rare NFs. The increased isolation and identification of these NFs requires the attention of clinicians to understand and appropriately treat these organisms. With inadequate clinical associations, lack of susceptibility breakpoints, diverse susceptibility profiles, and absence of definite therapy regimens, it's challenging to deal with these emerging, uncommon non-fermenters. This study also highlights the necessity to formulate explicit guidelines for the antibiotic susceptibility testing of such uncommon NFs. Further studies are essential to understand the significance of this organism in human ailments as a colonizer and/or infecting agent.

Contributor Statement

NT, BM conceived the idea and edited the manuscript. KZ, PG, VK undertook the literature search, data extraction, and drafted the report. KZ, PG, VK assisted in interpretation and critical revision of the report. All the authors contributed equally. All the authors have read and approved the manuscript.

Declaration of Interests

The authors declare that they have no conflicts of interest.

Role of Funding Source

None

Acknowledgment

None

References

1. Warren JW. (1996) Clinical presentations and epidemiology of urinary tract infection. In: Mobley HL, Warren JW, ed. Urinary tract infections molecular pathogenesis and clinical management. Washington DC: American Society for Microbiology Press: 3-27. Van der

2. Kaushal ML, Grover PS, Gupta ML. Non-fermenters in urinary tract infection. *J Assoc Physicians India*. 1998;46(9):798-800.
3. Gupta P, Zaman K, Mohan B, Taneja N. Elizabethkingia miricola: A rare non-fermenter causing urinary tract infection. *World J Clin Cases*. 2017;5(5):187-190.
4. Velden LBJ, de Jong AS, de Jong H, de Gier RPE, Rentenaar RJ. First report of a *Empedobacter falsenii* isolated from the urine of an infant with pyelonephritis. *Diagnostic Microbiology and Infectious Disease*. 2012;74(4):404-405.
5. Traglia GM, Dixon C, Chiem K, et al. Draft genome sequence of *Empedobacter* (formerly *Empedobacter*) *falsenii* comb. nov. Wf282, a strain isolated from a cervical neck abscess. *Genome Announc*. 2015;3(2):235-245.
6. Kampfer P, Avesani V, Janssens M, Charlier J, De Baere T, Vaneechoutte M. Description of *Empedobacter falsenii* gen. nov., sp. nov., to accommodate clinical isolates phenotypically resembling members of the genera *Chryseobacterium* and *Empedobacter*. *Int J Syst Evol Microbiol*. 2006;56(10):2323-2329.
7. Cesira Giordano, Margherita Falleni, Anna-Lisa Capria, Francesco Caracciolo, Mario Petrini, Band Simona Barninia. First report of *Empedobacter falsenii* genomovar 2 isolated from the respiratory tract of an immunosuppressed man. *ID Cases*. 2016;4:27-29.
8. Marchandin H, Michon AL, Jumas Bilak E. Atypical bacteria in the CF airways: diversity, clinical consequences, emergence and adaptation. In: Sriramulu D, ed. *Cystic fibrosis - renewed hopes through research. InTech*. 2012.
9. Xu Y, Rudkjobing VB, Simonsen O, et al. Bacterial diversity in suspected prosthetic joint infections: an exploratory study using 16S rRNA gene analysis. *FEMS Immunol Med Microbiol*. 2012;65(2):291-304.
10. Harris D, Pacheco A, Lindner AS. Detecting potential pathogens on hospital surfaces: an assessment of carpet tile flooring in the hospital patient environment. *Indoor Built Environ*. 2010;19(2):239-249.
11. Perkins SD, Angenent LT. Potential pathogenic bacteria in metalworking fluids and aerosols from a machining facility. *FEMS Microbiol Ecol*. 2010;74(3):643-54.