

Is Beta-Amyloid Accumulation a Cause or Consequence of Alzheimer's Disease?

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Abstract

Alzheimer's disease (AD) is a progressive neurodegenerative disorder that is characterized by the pathological hallmarks of extracellular beta-amyloid (A β) plaques and intraneuronal tau-containing neurofibrillary tangles in the brain. Intraneuronal accumulation of A β also plays a role to accelerate AD progression by promoting neurodegeneration. Additionally, AD is associated with the development of amyloid angiopathy (CAA), in which A β builds up on the walls of the cerebral arteries, which augments the development of cerebral vascular disease (CVD). Conversely, CVD promotes A β deposition and the development of AD by affecting the balance of A β production and clearance. However, it remains to be determined whether the accumulation of A β is a cause or consequence of AD. The interaction between AD and CVD is a topic of considerable current interest. Here, we discuss the role of CVD in A β accumulation and the development of AD to provide a new point of view that combination therapies that address the accompanying cerebral microvascular disease may potentiate the efficacy of emerging treatment for AD.

Keywords: Alzheimer's disease, Cerebral vascular disease, Beta-Amyloid, Blood-brain barrier

Alzheimer's disease (AD) is the most common form of dementia. There is no cure and current therapies are not effective in delaying progression. An estimated 5.4 million Americans of all ages have AD. One in eight patients older than 65 years and 50% of the population \geq 85 years of age are affected by this neurodegenerative disease [1-3]. The Medicare cost for the treatment of dementia and AD is \$159 billion annually and is projected to rise to \$511 billion by 2040 [1].

Pathological Characteristics of Alzheimer's Disease

AD is characterized by the accumulation of intracellular tau-containing neurofibrillary tangles and extracellular β -amyloid (A β) deposits in the brain. A β is formed by sequential cleavage of the amyloid precursor protein (APP) in the cell membrane. β -secretase (BACE) first removes the ectodomain of APP, and then γ -secretase cleaves the remaining C-terminal fragment of APP's transmembrane domain to liberate A β from the cellular membrane to the extracellular space. The A β protein aggregates into plaques that enhance the formation of neuro-fibrillary tangles [3-6]. These tangles promote the loss of neurons and synaptic density by enhancing inflammation, oxidative stress and the development of cerebral microvascular disease [7,8]. A β plaques also promote the loss of microglial cells and the senescence of neural stem/progenitor cells by affecting forebrain and hippocampal neurogenesis and eliciting a senescence response in associated astrocytes [9-11]. Recent studies have indicated that A β protein also increases the expression of p16 and accelerates the formation of post-mitotic neurons in the central nervous system of elderly mice, which is associated with increases in the severity of cognitive impairments and dementia [12].

Although AD is primarily associated with extracellular A β plaques and neurofibrillary tangles in the brain, recent studies have indicated that intra-neuronal accumulation of A β also accelerates AD progression by promoting degeneration and loss of neurons [13]. The levels of intracellular soluble A β protein have been found to correlate with the loss of neuronal synapses and cognitive impairment, whereas A β plaques showed little correlation [14]. A recent study showed that the protein PirB is a receptor for soluble A β . When the receptor is bound to soluble A β , it leads to synapse degradation by biochemically altering and increasing the activity of cofilin, an enzyme

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that degrades actin which is crucial to the integrity of neuronal synapses [15]. Another study using double transgenic mice with mutations in the APP and presenilins (PS1, PS2) genes, reported that the production of A β and the expression of p25 was elevated [13]. The rise in A β production causes an increase in intracellular calcium that activates calpain which in turn activates p25. P25 is thought to cause deregulation of cdk5, [16] a protein kinase involved in the development of the CNS. When cdk5 regulation is interrupted, the cytoskeleton is disturbed and this leads to neuron death by an unknown mechanism [16]. Moreover, ischemic or hypoxic conditions in the brain have also been found to up regulate the expression of enzymes that produce A β protein [14]. Ischemia of the brain causes an increase in the release of pro-inflammatory mediators, which in turn modify the metabolism pathway and increase the production of APP [17]. The increase in APP triggers an increase in β - and γ -secretases, which amplify the production of A β and leads to intraneuronal A β accumulation [17]. Therefore, vascular dysfunction and hypoxic conditions in the brain can have neurotoxic effects on synapses leading to neuronal destruction causing the cognitive impairments seen in AD [14].

It is well recognized that A β plaques are hallmark diagnostic biomarkers of AD [18]. However, current treatments and clinical trials targeting A β with several γ -secretase inhibitors, which have effectively reduced A β protein levels, have failed in clinical trials. Moreover, γ -secretase inhibitors appear to have adverse effects on cognition themselves, secondary to activation of Notch or other critical substrates [19-21]. The low efficacy of current therapeutics for the treatment of AD has led the community to reconsider the idea whether an increase in the production of A β is the primary underlying cause of AD. Hence, there is considerable interest to better understand the mechanisms leading to the accumulation of A β protein in the brain, and to better define under what conditions they become clinically significant.

Cerebral Vascular Disease and Alzheimer's Disease

There is increasing evidence that cerebral microvascular dysfunction plays an important role in the development of AD. Indeed, the interaction between cerebral vascular disease (CVD) and AD is a topic of considerable current interest [22-26]. However, it remains to be determined whether CVD in AD patients is secondary to the accumulation of A β , or if it serves as an initiating factor in the development of this neurodegenerative disease. Hypertension, diabetes, and hyperlipidemia, along with aging, are the primary risk factors for AD. They all are the leading risk factors for cerebral vascular disease, endothelial dysfunction and stroke. Almost all elderly patients with severe CVD exhibit substantial A β accumulation [20,27,28]. Conversely, infarcts and small vessel disease, which are the hallmarks of vascular dementia (VaD) [28,29], are found in >50% of brains collected from AD patients [23,24]. Based on this evidence, several investigators have suggested that AD may be a primary vascular disorder rather than secondary to A β deposition induced neurodegeneration [23-25].

An imbalance of A β production and clearance results in accumulation of A β surrounding cerebral vessels [30] and produces amyloid angiopathy (CAA) in up to 90% of the brains of AD patients, versus only about 30% in controls [23,28].

Most patients with AD exhibit small vascular disease (SVD) and ischemic parenchymal abnormalities [28]. SVD and cerebral infarcts, which are the hallmarks of vascular dementia, [28,29] are found in >50% of brains collected from AD patients [23,24]. The incidence of AD among elderly patients is 2-fold higher after ischemic stroke, which causes localized hypoxia in the brains [31]. APP levels are elevated, and β -secretase is activated following cerebral ischemia in animal models [32].

Micro-RNAs (miRNAs) also play an important role in hypoxia stress responses. MiR-124 was found to be inhibited from the early to the late stages of cerebral hypoxia. This was accompanied with upregulation of expression of BACE1 protein, and an increase in A β protein levels in the hippocampus in rat models of cerebral ischemia [33]. ApoE4 may also accelerate the production and accumulation of A β protein by impairing the transport of A β protein from the brain to the cerebral spinal fluid (CSF) across the blood-brain barrier (BBB) [31]. Under hypoxic conditions, autophagy-lysosome mediated proteolysis of ApoE is disturbed and the subsequent increase in ApoE4 protein enhances the accumulation of A β protein [34]. Hypoxia also activates inflammatory pathways and increases the formation of neurofibrillary tau tangles and A β plaques, as well as reduces A β degradation and the clearance of A β protein across the BBB [31,35].

Blood-brain Barrier and Alzheimer's Disease

The BBB is a highly specialized endothelial cell membrane that lines cerebral microvessels. It represents the interface between neural and circulating cells of the immune system. SVD of AD patients are associated with endothelial dysfunction. The expression of CD31 and CD34 is decreased in AD which leads to an impairment of transport of proteins across the BBB from the CSF back into the circulation [36,37]. Lipoprotein receptor-related protein 1 (LRP1) and the receptor for advanced glycation end products (RAGE) are expressed in the cerebral microvascular endothelial cells, and regulate the clearance of A β from the central nervous system (CNS). LRP1 mediates the efflux of A β from neurons and astrocytes to the extracellular fluid in the brain, whereas RAGE promotes the accumulation of A β protein from the blood into the brain [38,39]. Clinical studies have demonstrated that AD pathology correlates with lower LRP1 and higher RAGE levels, which promote the accumulation of A β in the brain parenchyma [35,39,40]. These results support the idea that the loss of the expression and function of these transporters mediating both efflux of proteins and endothelial dysfunction, allows BBB leakage and the movement of proteins from the circulation into the CNS, and contributes to A β accumulation in AD.

Hypertension and aging are among the primary risk factors for AD. They act synergistically to accelerate the progression of AD pathology. Loss of BBB integrity occurs in the brain in both experimental and genetic models of hypertension [41-43]. Elevated blood pressure in AD may increase BBB leakage and the diffusion of circulating proteins, cytokines and immune cells into the CNS that promote inflammation and activate microglia in AD [44]. Microglia normally plays a protective role early in the pathogenesis of AD by increasing the clearance of A β protein by secreting neprilysin (NEP) and insulin-degrading enzyme (IDE). An increase in anti-inflammatory cytokine IL-4 increases the

expression of the scavenger receptor CD36 and the A β degrading enzymes NEP and IDE in rat primary type 2 microglia [45]. However, in late stages of AD, over-activation of the microglia leads to inflammation and the production of pro-inflammatory cytokines, such as TNF- α , IL-1 β , and IFN- γ , that promotes the accumulation of A β , decreases A β clearance by cerebral vascular endothelial cells, increases the permeability of BBB and promotes neuroinflammation [37,44,46].

Cerebral Autoregulation and Alzheimer's Disease

Auto regulation of cerebral blood flow (CBF) is a vital homeostatic mechanism that maintains adequate blood flow to the brain despite fluctuations in pressure [47]. It protects the brain from increases in capillary pressure, which is associated with BBB leakage, cerebral edema and neurological damage [42,47-51]. Recent studies have revealed that auto regulation of CBF is impaired in elderly hypertensive, diabetic and AD animal models. Impaired cerebral autoregulation increases transmission of pressure to small vessels which contributes to BBB disruption, inflammation, glial activation, capillary loss, microhemorrhage, localized ischemia and neurodegeneration, which accelerate the cognitive deficits in both VaD and AD patients [52-55].

Conclusion

To date, no safe and effective treatments for AD have been emerged. The hypotheses of the underlying cause of neurodegenerative AD have been focusing on cholinergic deficiency, extracellular A β accumulation that forms plaques in the brain, and Tau protein abnormalities leading to neurofibrillary tangles. However, current treatments and clinical trials targeting these pathways, such as using acetylcholinesterase inhibitors and γ secretase inhibitors, have not been effective which has led the community to reconsider the alternatives [21,22,56-61]. In addition, γ -secretase inhibitors that are effective in reducing A β protein in animal models of AD have been linked to a higher incidence of skin cancers and increases in cognitive impairment in AD patients [14]. Anti-A β immunotherapy has emerged as one of the most promising therapeutic strategies for the treatment of AD via microglial recruitment, Fc γ receptor-mediated phagocytosis, and a peripheral sink mechanism. However, investigators recently reported that there was no cognitive benefit of immunotherapy in a co-morbidity mouse model which exhibits amyloid deposition comorbid with VaD (APP/PS1 HHcy) [62]. This model develops additive cognitive deficits and redistribution of amyloid favoring the deposition of cerebral amyloid CAA [62]. Moreover, even though anti-A β immunotherapy has been demonstrated to delay loss of cognition in preclinical animal model studies, the outcome of recent clinical trials have been disappointing and failed to meet primary efficacy end points.

Taken together, it appears that CVD and BBB dysfunction play an integral role in the development of AD by increasing the accumulation of A β protein. Hence, the development of new therapeutic strategies for the treatment of AD should take the associated CVD into consideration. It is our view that combination therapies that address the accompanying microvascular disease and that use drugs to provide strict control of associated hypertension and diabetes, along with disease modifying drugs used to control inflammation in rheumatoid arthritis, may potentiate the efficacy of the γ -secretase inhibitors and anti-A β immunotherapy for the treatment of AD.

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References

1. Hurd MD, Martorell P, Delavande A, Mullen MJ, Langa KM. Monetary Costs of Dementia in the United States. *N Engl J Med*. 2013;368:1326-1334.
2. Pleis JR, Lethbridge-Cejku M. Summary health statistics for U.S. adults: National Health Interview Survey, 2005. *Vital Health Stat 10*. 2006;(234):1-153.
3. Gotz J, Schild A, Hoerndli F, Pennanen L. Amyloid-induced neurofibrillary tangle formation in Alzheimer's disease: Insight from transgenic mouse and tissue-culture models. *Int J Dev Neurosci*. 2004;22(7):453-465.
4. Bolduc DM, Montagna DR, Seghers MC, Wolfe MS, Selkoe DJ. The amyloid-beta forming tripeptide cleavage mechanism of γ -secretase. *eLIFE*. 2016;5:1-21.
5. Steiner H, Capell A, Leimer U, Haass C. Genes and mechanisms involved in -amyloid generation and Alzheimer's disease. *Eur Arch Psychiatry Clin Neurosci*. 1999;249:266-270.
6. Seino Y, Kawarabayashi T, Wakasaya Y, et al. Amyloid β accelerates phosphorylation of tau and neurofibrillary tangle formation in an amyloid precursor protein and tau double-transgenic mouse model. *J Neurosci Res*. 2010;88(16):3547-3554.
7. Reiman EM. Attack on amyloid- β protein. *Nature*. 2016;537:36-37.
8. Lue LF, Brachova L, Civin HW, Rogers J. Inflammation, A β Deposition, and neurofibrillary tangle formation as correlates of Alzheimer's disease neurodegeneration. *J Neuropathol Exp Neurol*. 1996;55(10):1083-1088.
9. He N, Jin W-L, Lok KH, Wang Y, Yin M, Wang ZJ. Amyloid- β (1-42) oligomer accelerates senescence in adult hippocampal neural stem/progenitor cells via formylpeptide receptor 2. *Cell Death Dis*. 2013;4:e924.
10. Molofsky AV, Slutsky SG, Joseph NM, et al. Increasing p16INK4a expression decreases forebrain progenitors and neurogenesis during ageing. *Nature*. 2006;443:448-452.
11. Bhat R, Crowe EP, Bitto A, et al. Astrocyte senescence as a component of Alzheimer's disease. *PLoS One*. 2012;7:e45069.
12. Wei Z, Chen XC, Song Y, et al. Amyloid β protein aggravates neuronal senescence and cognitive deficits in 5XFAD mouse model of Alzheimer's disease. *Chin Med J (Engl)*. 2016;129(15):1835-1844.
13. Oakley H, Cole SL, Logan S, et al. Intraneuronal beta-amyloid aggregates, neurodegeneration, and neuron loss in transgenic mice with five familial Alzheimer's disease mutations: potential factors in amyloid plaque formation. *J Neurosci*. 2006;26(40):10129-10140.
14. Honjo K, Black SE, Verhoeff NP. Alzheimer's disease, cerebrovascular disease, and the beta-amyloid cascade. *Can J Neurol Sci*. 2012;39(6):712-728.
15. Kim T, Vidal GS, Djuricic M, et al. Human LirB2 is a beta-amyloid receptor and its murine homolog PirB regulates synaptic plasticity in an Alzheimer's model. *Science*. 2013;341(6152):1399-1404.
16. Cruz JC, Tsai LH. Cdk5 deregulation in the pathogenesis of Alzheimer's disease. *Trends Mol Med*. 2004;1(9):452-458.
17. Pluta R, Ułamek M, Jablonski M. Alzheimer's mechanisms in ischemic brain degeneration. *Anat Rec (Hoboken)*. 2009;292(12):1863-1881.
18. Prvulovic D, Hampel H. Amyloid β (A β) and phospho-tau (p-tau) as diagnostic biomarkers in Alzheimer's disease. *Clin Chem Lab Med*. 2011;49(3):367-374.
19. Coric V, van Dyck CH, Salloway S, et al. Safety and tolerability of the γ -Secretase inhibitor avagacestat in a phase 2 study of mild to moderate Alzheimer disease. *Arch Neurol*. 2012;69(11):1430-1440.

20. Farkas E, Luiten PG. Cerebral microvascular pathology in aging and Alzheimer's disease. *Prog Neurobiol*. 2011;64(6):575-611.
21. Bateman RJ, Siemers ER, Mawuenyega KG, et al. A gamma-secretase inhibitor decreases amyloid-beta production in the central nervous system. *Ann Neurol*. 2009;66(1):48-54.
22. Doody RS, Raman R, Farlow M, et al. A phase 3 trial of semagacestat for treatment of Alzheimer's disease. *N Engl J Med*. 2013;369:341-350.
23. Bell RD, Zlokovic BV. Neurovascular mechanisms and blood-brain barrier disorder in Alzheimer's disease. *Acta Neuropathol*. 2009;118(1):103-113.
24. Jellinger KA. Pathology and pathogenesis of vascular cognitive impairment—a critical update. *Front Aging Neurosci*. 2013;5:17.
25. De la Torre JC. Vascular basis of Alzheimer's pathogenesis. *Ann N Y Acad Sci*. 2002;977:196-215.
26. Attems J, Jellinger KA. The overlap between vascular disease and Alzheimer's disease - lessons from pathology. *BMC Med*. 2014;12:206.
27. De-Paula VJ, Radanovic M, Diniz BS, Forlenza OV. Alzheimer's disease. *Subcell Biochem*. 2012;65:329-352.
28. Love S, Miners JS. Cerebrovascular disease in aging and Alzheimer's disease. *Acta Neuropathol*. 2016;131(5):645-658.
29. Iadecola C. Neurovascular regulation in the normal brain and in Alzheimer's disease. *Nat Rev Neurosci*. 2004;5(5):347-360.
30. Zlokovic B V, Yamada S, Holtzman D, Ghiso J, Frangione B. Clearance of amyloid beta-peptide from brain: transport or metabolism? *Nat Med*. 2006;6(7):718-719.
31. Zhang X, Le W. Pathological role of hypoxia in Alzheimer's disease. *Exp Neurol*. 2010;223(2):299-303.
32. Brambillaa A, Lonati E, Milani C, et al. Ischemic conditions and β -secretase activation: The impact of membrane cholesterol enrichment as triggering factor in rat brain endothelial cells. *Int J Biochem Cell Biol*. 2015;69:95-104.
33. Zhang X, Huang X, Fang C, et al. miR-124 Regulates the Expression of BACE1 in the Hippocampus Under Chronic Cerebral Hypoperfusion. *Mol Neurobiol*. 2016.
34. Zhou W, Scott SA, Shelton SB, Crutcher KA. Cathepsin D-mediated proteolysis of apolipoprotein E: Possible role in Alzheimer's disease. *Neuroscience*. 2006;143(3):689-701.
35. Bell RD, Deane R, Chow N, et al. SRF and myocardin regulate LRP-mediated amyloid-beta clearance in brain vascular cells. *Nat Cell Biol*. 2009;11(2):143-153.
36. Kalaria RN. Neuropathological diagnosis of vascular cognitive impairment and vascular dementia with implications for Alzheimer's disease. *Acta Neuropathol*. 2016;131(5):659-685.
37. Zenaro E, Piacentino G, Constantin G. The blood-brain barrier in Alzheimer's disease. *Neurobiol Dis*. 2016.
38. Deane R, Wu Z, Zlokovic BV. RAGE (Yin) Versus LRP (Yang) Balance Regulates Alzheimer Amyloid -Peptide Clearance Through Transport Across the Blood-Brain Barrier. *Stroke*. 2004;35:2628-2632.
39. Deane R, Singh I, Sagare AP, et al. A multimodal RAGE-specific inhibitor reduces amyloid β -mediated brain disorder in a mouse model of Alzheimer disease. *J Clin Invest*. 2012;122(4):1377-1392.
40. Donahue JE, Flaherty SL, Johanson CE, et al. RAGE, LRP-1, and amyloid-beta protein in Alzheimer's disease. *Acta Neuropathol*. 2006;122(4):405-415.
41. Meissner A. Hypertension and the Brain: A Risk Factor for More Than Heart Disease. *Cerebrovasc Dis*. 2016;42:255-262.
42. Fan F, Geurts AM, Pabbidi MR, et al. Zinc-finger nuclease knockout of dual-specificity protein phosphatase-5 enhances the myogenic response and autoregulation of cerebral blood flow in FHH.1BN rats. *PLoS one*. 2014;9(11):e112878.
43. Winklewski PJ, Radkowski M, Demkow U. Neuroinflammatory mechanisms of hypertension: potential therapeutic implications. *Curr Opin Nephrol Hypertens*. 2016;25(5):410-416.
44. Sumbria RK, Grigoryan MM, Vasilevko V, et al. A murine model of inflammation-induced cerebral microbleeds. *J Neuroinflammation*. 2016;13(1):218.
45. Shimizu E, Kawahara K, Kajizono M, Sawada M, Nakayama H. IL-4-induced selective clearance of oligomeric beta-amyloid peptide (1-42) by rat primary type 2 microglia. *J Immunol*. 2008;181(9):6503-6513.
46. Wang Y, Jin S, Sonobe Y, et al. Interleukin-1 β induces blood-brain barrier disruption by downregulating sonic hedgehog in astrocytes. *PLoS One*. 2014;9(10):1-8.
47. Faraco G, Iadecola C. Hypertension: a harbinger of stroke and dementia. *Hypertension*. 2013;62(5):810-817.
48. Gorelick PB, Pantoni L. Advances in vascular cognitive impairment. *Stroke*. 2013;44(2):307-308.
49. Iadecola C. Hypertension and dementia. *Hypertension*. 2014;64(1):3-5.
50. Fan F, Geurts AM, Murphy SR, Pabbidi MR, Jacob HJ, Roman RJ. Impaired myogenic response and autoregulation of cerebral blood flow is rescued in CYP4A1 transgenic Dahl salt-sensitive rat. *Am J Physiol Regul Integr Comp Physiol*. 2014;308(5):R379-390.
51. Fan F, Ge Y, Lv W, et al. Molecular mechanisms and cell signaling of 20-hydroxyeicosatetraenoic acid in vascular pathophysiology. *Front Biosci (Landmark Ed)*. 2016;21:1427-1463.
52. Faraci FM, Baumbach GL, Heistad DD. Cerebral circulation: humoral regulation and effects of chronic hypertension. *J Am Soc Nephrol*. 1990;1(1):53-57.
53. Harder DR, Narayanan J, Gebremedhin D. Pressure-induced myogenic tone and role of 20-HETE in mediating autoregulation of cerebral blood flow. *Am J Physiol Heart Circ Physiol*. 2011;300(5):H1557-1565.
54. Paulson OB, Strandgaard S, Edvinsson L. Cerebral autoregulation. *Cerebrovasc Brain Metab Rev*. 1990;2(2):161-192.
55. Lammie GA. Hypertensive cerebral small vessel disease and stroke. *Brain Pathol*. 2002;12(3):358-370.
56. Randall J, Bateman, Ling Y, Munsell, Xianghong Chen, David M. Holtzman, Kevin E. Yarasheski. Stable isotope labeling tandem mass spectrometry (SILT) to quantify protein production and clearance rates. *Journal of the American Society for Mass Spectrometry*. 2007;18(6):997-1006.
57. Siemers ER, Dean RA, Friedrich S, et al. Safety, tolerability, and effects on plasma and cerebrospinal fluid amyloid-beta after inhibition of gamma-secretase. *Clin Neuropharmacol*. 2007;30(6):317-325.
58. Siemers ER, Quinn JF, Kaye J, et al. Effects of a gamma-secretase inhibitor in a randomized study of patients with Alzheimer disease. *Neurology*. 2006;66(4):602-604.
59. Fleisher AS, Raman R, Siemers ER, et al. Phase 2 safety trial targeting amyloid beta production with a gamma-secretase inhibitor in Alzheimer disease. *Arch Neurol*. 2008;65(8):1031-1038.
60. Doody RS, Aisen PS, Iwatsubo T. Semagacestat for treatment of Alzheimer's disease. *N Engl J Med*. 2013;369:1660-1661.
61. Doody RS, Raman R, Farlow M, et al. Semagacestat Study, G: A phase 3 trial of semagacestat for treatment of Alzheimer's disease. *N Engl J Med*. 2013;369:341-350.
62. Weekman EM, Sudduth TL, Caverly CN, et al. Reduced Efficacy of Anti-A β Immunotherapy in a Mouse Model of Amyloid Deposition and Vascular Cognitive Impairment Comorbidity. *J Neurosci*. 2016;36(38):9896-9907.